

## Recommendations for Childhood Cancer Screening and Surveillance in DNA Repair Disorders



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### Abstract

DNA repair syndromes are heterogeneous disorders caused by pathogenic variants in genes encoding proteins key in DNA replication and/or the cellular response to DNA damage. The majority of these syndromes are inherited in an autosomal-recessive manner, but autosomal-dominant and X-linked recessive disorders also exist. The clinical features of patients with DNA repair syndromes are highly varied and dependent on the underlying genetic cause. Notably, all patients have elevated risks of syndrome-associated cancers, and many of these cancers present in childhood. Although it is clear that the risk of cancer is increased, there are limited data defining the true incidence of cancer and almost no evidence-based approaches to cancer surveillance in patients with DNA repair disorders. This article is the product of the October 2016 AACR Childhood Cancer Predisposition Workshop, which brought together experts from

around the world to discuss and develop cancer surveillance guidelines for children with cancer-prone disorders. Herein, we focus on the more common of the rare DNA repair disorders: ataxia telangiectasia, Bloom syndrome, Fanconi anemia, dyskeratosis congenita, Nijmegen breakage syndrome, Rothmund–Thomson syndrome, and Xeroderma pigmentosum. Dedicated syndrome registries and a combination of basic science and clinical research have led to important insights into the underlying biology of these disorders. Given the rarity of these disorders, it is recommended that centralized centers of excellence be involved directly or through consultation in caring for patients with heritable DNA repair syndromes. *Clin Cancer Res*; 23(11); e23–e31. ©2017 AACR.

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### Introduction

Germline pathogenic variants (i.e., mutations) in key components of DNA repair and telomere biology result in a spectrum of heritable disorders usually associated with characteristic physical findings and an elevated risk of specific cancers. In many instances, the DNA repair disorders are diagnosed in childhood, but some, particularly those caused by aberrant telomere biology, may manifest later in life. Dedicated syndrome registries, basic science, and clinical research have provided insights into the treatment and management for

individuals with these rare disorders of aberrant DNA repair mechanisms.

This article originates from the October 2016 AACR Childhood Cancer Predisposition Workshop, which focused on reviewing pediatric cancer surveillance guidelines for children with hereditary risk of cancer. Limited data exist to define the true incidence of cancer in the DNA repair disorders, and almost no evidence-based approaches exist to evaluate cancer surveillance in patients with DNA repair disorders. As the comprehensive review of all inherited disorders of DNA repair is beyond the scope of this article and has been done elsewhere, we reviewed the primary clinical manifestations and associated malignancies of these disorders, and provide information on family support groups and/or patient registries as a starting point for clinical management and the future development of evidence-based guidelines (Tables 1 and 2; refs. 1–8). All patients and/or their families are encouraged to promptly report to health care professionals any changes in their health, and physicians should have a low index of suspicion for malignancy in patients with DNA repair disorders. In addition to contacting centers of excellence for these rare disorders, clinicians are referred to <https://clinicaltrials.gov> to help identify ongoing clinical trials for these disorders.

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### Ataxia Telangiectasia

#### Genetic summary

Ataxia telangiectasia (AT; OMIM #208900) is an autosomal-recessive (AR) disorder presenting in childhood due to biallelic pathogenic variants in the *ATM* (ataxia-telangiectasia mutated)

**Table 1.** Genetic features of DNA repair and telomere biology disorders

Disorder	Biological pathway	Inheritance: gene(s)
Ataxia telangiectasia	DNA repair checkpoints	AR: <i>ATM</i>
Bloom syndrome	Homologous recombination	AR: <i>BLM</i>
Dyskeratosis congenita	Telomere biology	XLR: <i>DKC1</i> AD: <i>TERC, TERT, TINF2, RTEL1, PARN, NAF1</i> AR: <i>NOPI0, NHP2, TERT, RTEL1, PARN, CTC1, STN1, POT1, WRAP53, ACD</i>
Fanconi anemia	DNA damage response, especially interstrand cross-link repair	AR: <i>FANCA, FANCC, FANCD1/BRCA2, FANCD2, FANCE, FANCF, FANCG, FANCI, FANCL, FANCD1/BRIP1/BACH1, FANCL, FANCM, FANCN/PALB2, FANCO/RAD51C, FANCP/SLX4, FANCP/XP/ERCC4, FANCR/RAD51, FANCES/BRCA1, FANCT/UBE2T, FANCU/XRCC2, REV7/MAD2L2</i> XLR: <i>FANCB</i>
Nijmegen breakage syndrome	DNA double-stranded break repair	AR: <i>NBN</i>
Rothmund-Thomson syndrome	DNA replication/repair helicase	AR: <i>RECQL4</i>
Xeroderma pigmentosum	Nucleotide excision repair	AR: <i>DBP2, ERCC1, ERCC2, ERCC3, ERCC4, ERCC5, POLH, XPA, or XPC</i>

Abbreviations: AD, autosomal dominant; AR, autosomal recessive; XLR, X-linked recessive.

gene, which encodes a protein belonging to the phosphatidylinositol-3 kinase (PI3) protein family. The incidence of A-T is estimated between 1:40,000 and 1:100,000 people (9). The ATM protein is a cell-cycle checkpoint kinase that functions as a regulator of multiple proteins, including tumor suppressor proteins p53, BRCA1, CHEK2, and NBS1 (10). Pathogenic variants in *ATM* typically decrease the expression and/or function of ATM and prevent cells from responding correctly to DNA damage, which allows breaks in DNA strands to accumulate and contributes to genomic instability and/or cell death. This results in increased sensitivity to ionizing radiation in cells of patients with A-T (11). Patients with A-T typically develop progressive cerebellar ataxia between one and four years of age. Conjunctival telangiectasias, oculomotor apraxia, choreoathetosis, and immunodeficiency are often also present (1). The progressive neurologic symptoms are thought to be due to aberrant DNA repair and neuronal cell death, with most children with A-T wheelchair bound by the teen years. Malignancy is reported to develop in up to 40% of patients with A-T, and is typically non-Hodgkin lymphoma and acute lymphoid leukemia (12).

#### Cancer screening/surveillance/management protocols

Children with A-T are often diagnosed by a variety of different methods, including abnormal newborn screening for reduced T-cell receptor excision circle levels. Other laboratory abnormalities that can be detected in children suspected of having A-T include increased alpha-feto protein (AFP) levels; reduced IgA, IgE, and IgG2 levels; poor antibody response to pneumococcal polysaccharide vaccines; abnormal peripheral blood karyotype analysis including presence of a 7;14 translocation (in 5%–15% of patients); and cerebellar hypoplasia on MRI (1). Increased lymphocyte sensitivity to ionizing radiation is also present.

Patients with A-T require multidisciplinary care including referrals to (i) neurology for progressive cerebellar ataxia, ocular apraxia, and choreoathetosis; (ii) immunology/hematopoietic cell transplant (HCT) for the management of immunodeficiency; (iii) pulmonology for recurrent infections, pulmonary function evaluation, and restrictive lung disease; (iv) gastroenterology (GI) for swallow evaluation and nutrition; and (v) oncology for leukemia, lymphoma, and solid tumor risks.

Evidence-based standards for cancer screening do not exist for patients with A-T, particularly in childhood. Annual physical exam, complete blood count (CBC), and complete metabolic

profile including lactate dehydrogenase should be considered. As described in another article in this series, considerable debate exists on whether early diagnosis of acute leukemia improves survival (13). It is important for parents and providers to keep in mind that patients with A-T are sensitive to ionizing radiation and X-rays, and, thus, their use should be limited accordingly. Treatment regimens of any incident cancer should be adjusted given the increased risk of treatment related toxicity in children with A-T. Providers and patients should coordinate both acute and chronic care with centers focusing on A-T. The A-T Children's Project, <http://www.atcp.org>, has information for families and physicians.

Individuals who are heterozygous for a single pathogenic *ATM* variant have increased risk of adult onset breast, prostate, and pancreatic cancer (14–16). This also has implications for cancer screening in affected parents of children with A-T. The adult onset cancer risk in *ATM* carriers continues to be extensively studied but remains beyond the scope of this article.

## Nijmegen Breakage Syndrome

#### Genetic summary

Nijmegen breakage syndrome (NBS; OMIM #251260) is an AR disorder presenting in childhood due to biallelic pathogenic variants in *nibrin*, encoded by the *NBN* gene. *Nibrin* belongs to the MRE11/RAD50 double-stranded break repair complex. Patients with NBS are characterized by microcephaly, microgenia (small deformed chin), immunodeficiency, and "bird-like" facies (17). NBS is estimated to affect one in 100,000 newborns worldwide, but is thought to be more common in Slavic populations of Eastern Europe (18). Approximately 40% of affected individuals develop malignancies before age 20. T-cell and B-cell lymphomas are the most common NBS-associated malignancies; medulloblastoma, glioma, and rhabdomyosarcoma have also been reported (18).

Laboratory evaluation for NBS shows some similar features as described for children with A-T, including reduced CD3<sup>+</sup> and CD4<sup>+</sup> T cells, IgA deficiency (20% of patients), IgG2 and IgG4 deficiency (with normal serum IgG), increased frequency of CD45RO<sup>+</sup> T cells and simultaneous decrease in naïve CD45RA<sup>+</sup> T-cells (rare), and the same structural aberrations of chromosomes 7 and 14 in cultured lymphocytes as seen in AT. Response to testing of ionizing radiation sensitivity in lymphocytes will be abnormal and demonstrate increased sensitivity. Germline

**Table 2.** Diagnosis, associated malignancies, and management recommendations for DNA repair and telomere biology disorders

	<b>Diagnostic testing and other biomarkers</b>	<b>Associated malignancies</b>	<b>Management recommendations</b>
Ataxia telangiectasia	Genetic testing: newborn screening: reduced T-cell receptor excision circle levels Elevated alpha fetal protein Karyotype: 7:14 chromosomal translocation Immunoblotting (research) Chromosome breakage studies for radiation sensitivity (research)	Lymphoma, ALL, ovarian cancer, breast cancer, gastric cancer, melanoma, leiomyomas, sarcomas	Hematology-oncology: history/physical, annual CBCs, metabolic profile and lactate dehydrogenase and avoid excessive radiation Immunology: monitoring of immunoglobulin levels per immunologist recommendation Dermatology: annual skin exam Pulmonary: baseline and as needed pulmonary function tests Gastroenterology/nutrition: baseline and as needed swallowing function and nutritional management Endocrine: annual diabetes screen Neurology: supportive medications Orthopedics: annual scoliosis evaluation Dental: biannual exam
Bloom syndrome	Genetic testing: chromosome breakage with DEB and/or MMC abnormal sister chromatid exchange (research)	Lymphoma, AML, ALL, sarcoma, genital and urinary tract carcinoma, medulloblastoma, retinoblastoma	Hematology-oncology: CBCs every 3–4 months, avoidance of radiation, breast MRI/ultrasound starting at 18 years of age, annual colonoscopy starting at age 15 years, renal ultrasound examination at diagnosis and every 3 months through age 8 years to assess for Wilms tumor, HPV vaccine per AAP guidelines Dermatology: annual skin exam, limit sun exposure Pulmonary: pulmonary function tests Gastroenterology/nutrition: baseline and as needed swallowing function evaluation and nutritional management Endocrine: annual fasting blood sugar and TSH level Orthopedics: annual scoliosis evaluation Dental: biannual exam
Dyskeratosis congenita (telomere syndromes)	Genetic testing: telomere length measurement of leukocyte subsets using flow cytometry with <i>in situ</i> hybridization	Head and neck SCC, AML, MDS, anogenital cancer	Hematology-oncology: annual CBCs, ~annual bone marrow evaluations based on clinical features, early referral to HCT center, HPV vaccine, annual ENT evaluation beginning in adolescence Immunodeficiency: monitoring of immunoglobulin levels per immunologist recommendation Dermatology: annual skin exam Pulmonary: baseline pulmonary function tests with routine follow-up Gastroenterology/nutrition: liver function tests at least annually, more frequent if on androgens Endocrine: annual diabetes screen Neurology: MRI assessment for cerebellar hypoplasia at diagnosis, early intervention for developmental delay Ophthalmology: annual examination, monitoring and early management of lacrimal duct stenosis Orthopedics: evaluation of hip and shoulder avascular necrosis based on symptoms Dental: biannual exam ENT: baseline hearing evaluation Cardiology: baseline evaluation for AVMs and cardiac malformations Genitourinary: baseline assessment for genitourinary anomalies
Fanconi anemia	Genetic testing: chromosomal breakage with DEB and/or MMC	Head and neck SCC, AML, MDS, anogenital cancer	Hematology-oncology: annual CBCs, ~annual bone marrow evaluations based on clinical features, early referral to HCT center, HPV vaccine, annual ENT evaluation beginning in adolescence Immunology: monitoring of immunoglobulin levels per immunologist recommendation Dermatology: annual skin exam Pulmonary: baseline pulmonary function tests with follow-up as needed Gastroenterology/nutrition: annual liver function tests, more frequent if on androgens Endocrine: annual diabetes screen, follow growth trajectory Orthopedics: assessment of radial ray anomalies and management if needed Genitourinary: baseline evaluation for renal malformations Cardiology: baseline evaluation for heart malformations ENT: annual hearing evaluation, annual cancer screening starting in teenage years Dental: biannual exam

(Continued on the following page)

**Table 2.** Diagnosis, associated malignancies, and management recommendations for DNA repair and telomere biology disorders (Cont'd)

	<b>Diagnostic testing and other biomarkers</b>	<b>Associated malignancies</b>	<b>Management recommendations</b>
Nijmegen breakage syndrome	Genetic testing: chromosomal breakage with DEB and/or MMC Chromosomal instability involving chromosomes 7 and 14 in PHA-stimulated lymphocytes Immunoblotting (research)	Lymphoma, medulloblastoma, glioma, rhabdomyosarcoma	Hematology-oncology: history/physical, annual CBCs, metabolic profile and lactate dehydrogenase and avoid excessive radiation, HPV vaccine per AAP guidelines Dermatology: annual skin examinations Pulmonary: baseline pulmonary function tests with follow-up as needed, aggressive treatment of recurrent infections Gastroenterology/nutrition: baseline and as needed swallowing function evaluation and nutritional management Endocrine: monitor growth, assess females for ovarian failure Neurology: developmental assessment and early intervention if needed Ophthalmology: annual examination Orthopedics: baseline assessment for anomalies and as needed Dental: biannual exam
Rothmund-Thomson syndrome	Genetic testing	Osteosarcoma, basal cell carcinoma, skin SCC	Oncology: avoid ionizing radiation, consider imaging for osteosarcoma risk, HPV vaccine per AAP guidelines Dermatology: avoid excessive UV; use sunscreen annual exam and early treatment of lesions Ophthalmology: annual evaluation and cataract treatment as needed Endocrine: management for osteopenia Orthopedics: baseline skeletal survey Dental: biannual evaluation with proper care for hypoplastic teeth, enamel defects Oncology: beginning at diagnosis; avoid excessive sunlight and ionizing radiation; early identification and treatment of skin lesions; exam for ocular and ENT neoplasms every 6–12 months Dermatology: thorough skin evaluation every 3 months Gastroenterology/nutrition: evaluate swallowing function, nutritional support as needed Ophthalmology: exam every 6–12 months Neurology: evaluation for developmental delay or progressive neurologic changes Orthopedics: annual scoliosis evaluation ENT: baseline hearing evaluation and as needed cancer screening every 6–12 months
Xeroderma pigmentosa	Genetic testing	Melanoma, basal cell carcinoma, skin SCC, leukemia, brain and spinal cord tumors	

Abbreviations: AAP, American Academy of Pediatrics; ALL, acute lymphocytic leukemia; AML, acute myeloid leukemia; AML, acute myeloid leukemia; AVM, arteriovenous malformations; CBC, complete blood count; DEB, diepoxbutane; ENT, ear, nose and throat; HPV, human papillomavirus; IVIg, intravenous immunoglobulin; MDS, myelodysplastic syndrome; MMC, mitomycin C; PHA, phytohemagglutinin; SCC, squamous cell carcinoma.

genetic testing reveals loss-of-function mutations in *NBN*, with the Slavic founder mutation being the most common (6, 18).

#### Cancer screening/surveillance/management protocols

Patients with NBS require multidisciplinary care beginning at diagnosis, which is tailored to each patient's specific needs. Patients should be evaluated by an immunologist and for management of immunodeficiency, undergo monitoring by a pulmonologist for recurrent infections, and be followed by endocrinology and nutrition evaluations for growth deficiency and by oncology for leukemia, lymphoma, and solid tumor risks. Annual CBC is indicated or when symptomatic to assess for hematologic disease. Patients with NBS require often intravenous immunoglobulin therapy for immunodeficiency (19–22). As described for patients with A-T, children with NBS demonstrate increased sensitivity to ionizing radiation and may require tailored treatment regimens for any malignancy that develops (6).

Heterozygous carriers of pathogenic variants in *NBN* are at risk for adult onset breast and prostate cancer (15, 16, 23).

## Bloom Syndrome

### Genetic summary

Bloom syndrome (OMIM #210900) is an AR disorder resulting from biallelic pathogenic variants in the *BLM* gene encoding the BLM DNA helicase, a member of the RECQ family and sometimes referred to as *BLM* (24, 25). RECQ helicase enzymes attach and unwind the DNA double helix. BLM maintains genomic stability during the DNA copying process by limiting sister chromatid exchange. Cells from patients with Bloom syndrome with absent BLM activity demonstrate a 10 times higher rate of sister chromatid exchange.

Only a few hundred individuals with Bloom syndrome have been described, and approximately one third are of Ashkenazi Jewish descent due to a founder allele (26–28). The classic Bloom syndrome characteristics include pre- and postnatal growth deficiency, short stature, sun sensitivity, gastroesophageal reflux, recurrent infections, decreased fertility in males, insulin resistance, and cancer predisposition (4). A total of 212 cancers in 136 patients have been described in the Bloom Syndrome Registry (4). Cancers diagnosed during the pediatric period include gastrointestinal, genital and urinary tract carcinoma, lymphoma, acute lymphoblastic leukemia, acute myeloid leukemia (AML), sarcoma, Wilms tumor, medulloblastoma, and retinoblastoma (29). Multiple cancers occur commonly and with a distribution that is similar to cancer that is seen in the general population but with an earlier onset.

### Cancer screening/surveillance/management protocols

There is no established cancer screening protocol for patients with Bloom syndrome, and the risk for cancer at multiple sites presents a surveillance challenge. Patients and their families should be aware of the signs and symptoms of leukemia and lymphoma, the most commonly encountered malignancies in Bloom syndrome, and patients should be evaluated promptly when recognized. The second most common cancer type is colorectal cancer, with the earliest occurrence being at the age of 16 years (28). A reasonable approach to screening includes annual colonoscopy with fecal immunochemical testing every 6 months, beginning at age 15 years. Breast cancer was diagnosed in 17 women in the Bloom Syndrome Registry at a median age of 35.8 years (range 18–48). On the basis of this information, annual

breast MRI scans beginning at age 18 years is a reasonable surveillance strategy. Additional screening recommendations are provided in Table 2. For the remaining cancers, patients and their families should be aware of the common but nonspecific signs of cancer including unintentional weight loss, unexplained fever, fatigue, changes in bowel or bladder habits, and persistent and unexplained pain. When imaging is used for diagnostic evaluation, ultrasonography and MRI scan are preferred over radiographs or CT scans because of the presumed increased risk for cancer from ionizing radiation. The Bloom Syndrome Registry has information on various aspects of Bloom syndrome patient care (<http://weill.cornell.edu/bsr/>) as does the Bloom Syndrome Association (<http://www.bloomssyndromeassociation.org>). Clinicians should consult with experts in Bloom syndrome on the specific areas of clinical management. An international RECQ disorders meeting (RECQ2016) resulted in a recent plan to further develop Bloom syndrome management guidelines.

## Rothmund–Thomson Syndrome

### Genetic summary

Rothmund–Thomson syndrome (RTS; OMIM #268400) is a rare disorder with only a few hundred patients described in the literature (30, 31). Type 2 RTS is associated with an increased cancer risk results due to biallelic pathogenic variants in the *RECQL4* DNA helicase (8). The *RECQL4* protein, similar to BLM, belongs to the RECQ DNA helicase family. It is a multifunctional protein that participates in several cellular processes, including DNA replication, DNA damage repair, maintenance of telomeres, and mitochondrial DNA integrity (32, 33). A proportion of individuals with a clinical diagnosis of RTS (based on poikiloderma) do not have an identifiable *RECQL4* pathogenic variant (referred to as RTS type 1) and do not appear to have an increased risk of cancer. The gene for RTS type 1 has not been identified.

Patients with RTS have the characteristic skin finding of poikiloderma (hyper- and hypopigmentation, atrophy, and telangiectasias) that starts in infancy and persists throughout life (34–36). They may also have sparse hair, hyperkeratosis, small stature, skeletal defects including osteoporosis, dental anomalies, and cataracts (36–38). Patients with RTS develop osteosarcoma at an earlier age than the general population (median age 10 years), so any screening could be limited to the first two decades of life (39). The incidence of osteosarcoma in patients with no truncating mutations was 0.00 per year (100 person-years of observation), and the incidence of OS in patients with one or two truncating mutations was 0.05 per year (230 person-years of observation;  $P = 0.037$ ) using the two-sided log-rank test (40). A smaller number of patients have been described with basal cell carcinoma and skin squamous cell carcinoma (SCC; refs. 40, 41). A specific allele of *RECQL4* associated with a related disorder (RAPADILINO syndrome) is associated with an increased risk of lymphoma (42). Hematologic abnormalities, such as bone marrow failure (BMF), myelodysplastic syndrome (MDS), lymphoma, and leukemia have also been reported (36, 43–47).

### Cancer screening/surveillance/management protocols

Patients with RTS require multidisciplinary care including evaluations by (i) genetics for counseling about cancer risk, (ii) dermatology for annual skin exam and skin care, (iii) ophthalmology for cataract screening and management, and (iv) dentistry for routine care. Patients are cautioned to avoid excessive radiation (UV or IR) exposure, employ sensible sun protection, and

monitor skin for lesions. Retinoids may be used to manage hyperkeratosis, and pulsed laser therapy may be used to improve cosmesis of telangiectasias (48, 49). RTS patients with pathogenic variants in *RECQL4* are recommended to have a skeletal survey before the age of 5 years to identify any underlying skeletal abnormalities; they should receive counseling about the risk of osteosarcoma and be aware of signs and symptoms of osteosarcoma. Should these occur, they should seek immediate medical attention. Any new imaging of affected areas (e.g., X-rays) can be compared with the baseline skeletal survey to determine whether further workup is warranted. The benefit of routine screening for osteosarcoma has not yet been determined. Factors to consider include timing, length, modality (plain radiographs vs. MRI), and the cost of screening.

The RTS Foundation, <http://www.rtsplace.org/>, has patient-centered information.

## Dyskeratosis Congenita

### Genetic summary

Dyskeratosis congenita (OMIMs: #127550, #30500, #615190, #613987, #613989) is a telomere biology disorder (TBD) characterized by nail dystrophy, lacy skin pigmentation, and oral leukoplakia (7, 50). Dyskeratosis congenita is caused by pathogenic variants in genes important in stability and maintenance of telomeres, the nucleoprotein complex essential for chromosomal stability. The mode of inheritance depends on the gene and is X-linked for *DKC1*; autosomal dominant (AD) for *TERC* or *TINF2*; AR for *CTC1*, *NHP2*, *NOP10*, *PARN*, or *WRAP53*; and either AD or AR for *ACD*, *RTEL1*, or *TERT*. The prevalence of dyskeratosis congenita in the general population is unknown. Diagnosis is made by the presence of telomeres less than the first percentile for age measured by flow cytometry with FISH to measure telomere length in white blood cells (51). Patients with dyskeratosis congenita are at an increased risk of MDS; BMF; leukemia; cancers of the head and neck and genitourinary system; as well as pulmonary fibrosis, emphysema, and liver fibrosis/cirrhosis (51–53).

### Cancer screening/surveillance/management protocols

Diagnosis and clinical care guidelines for patients with dyskeratosis congenita were recently published (<https://www.dcoutreach.org/guidelines>). A bone marrow aspirate and biopsy are recommended after diagnosis of dyskeratosis congenita to establish a baseline. Because of the risk of developing MDS, CBCs and bone marrow evaluation should be performed at least annually but more often if clinically indicated due to signs, symptoms, or abnormal CBCs consistent with the familial leukemia report in this same series. Patients with dyskeratosis congenita on androgen therapy for cytopenias should undergo biannual hepatic ultrasounds and liver function tests every 3 months due to a potential risk of hepatic tumors and elevation of liver enzymes. Patients with dyskeratosis congenita are at high risk of head and neck squamous cell cancer (HNSCC) and, thus, should perform monthly oral self-examinations, have biannual dental examinations, and have annual HNSCC evaluation by an otolaryngologist experienced in oral cancer, beginning by age 16 years. An annual gynecologic examination is recommended for women. In addition, recommendations for nonmalignant sequelae include baseline pulmonary function tests when the patient is old enough to perform them with follow-up testing tailored to the individual

patient's needs. A baseline endocrine evaluation for growth is recommended and should be performed at least annually for patients on androgen therapy. Additional evaluation with developmental pediatricians and neurologists may be required for developmental delays (5, 54).

Additional resources for patients with dyskeratosis congenita are available at <https://www.dcoutreach.org>.

## Fanconi Anemia

### Genetic summary

Fanconi anemia (OMIMs: #134600, #227650, #600901, #609054, #605724, #613951, #610832, #614083, #614082, #609053) is a primarily AR disorder with at least 20 associated DNA repair genes. Pathogenic variants in one X-linked recessive gene, *FANCB*, and one AD gene, *FANCR* (*RAD51*), have been reported (3, 55–58). Fanconi anemia proteins function to maintain genomic stability by repairing DNA interstrand cross-links (ICL) and by interacting with other DNA damage response pathways (59, 60). The screening diagnostic test for Fanconi anemia involves chromosomal breakage assessment after exposure of T cells to diepoxybutane (DEB) or mitomycin C (MMC; refs. 61, 62).

Although the most common congenital anomalies include short stature, thumb or radii abnormalities, dysmorphic features, skeletal abnormalities, and genitourinary malformations, up to one third of patients will have no physical anomalies (63). Approximately 40% of patients with Fanconi anemia develop severe BMF by age 20 years and one half of all patients with Fanconi anemia develop BMF by the age of 50 years. The risks of solid tumors, including HNSCC, or AML by age 50 years in Fanconi anemia are estimated at 30% and 10%, respectively (52, 64–67). The success of HCT for BMF has led to improved survival in patients with Fanconi anemia but a possible increase in the incidence of HNSCC, kidney and liver tumors, brain tumors, breast cancers, and other tumor types (67, 68).

### Cancer screening/surveillance/management protocols

The guidelines for diagnosis and management of Fanconi anemia can be found at <http://fanconi.org/index.php/publications/guidelines>. A CBC and bone marrow aspirate and biopsy are recommended at diagnosis. The bone marrow evaluation should then be repeated annually. The CBC should be monitored more frequently to allow for proactive monitoring for progressive cytopenias and MDS. From the time of diagnosis, patients with Fanconi anemia should perform monthly oral self-examinations (or with parents' assistance) and have a biannual dental examination (general inspection exam without X-rays unless specific indication) and annual HNSCC evaluation by an otolaryngologist beginning in early adolescence. An annual gynecologic examination is recommended starting in adolescence, and the HPV vaccine should be administered per the American Academy of Pediatrics (AAP) vaccination schedule for both boys and girls. Clinical management of patients with Fanconi anemia does not include standard myeloablative dosing, as these lower dose regimens are designed to be myeloablative in the setting of Fanconi anemia; alternatively, androgen therapy may be tried for Fanconi anemia patients with BMF, as well as cancer-specific therapy with avoidance of DNA-damaging agents and supportive care for other complications (3).

Parents of children with the more common Fanconi anemia subtypes, *FANCA*, *FANCC*, and *FANCG*, do not appear to have

an increased cancer risk. However, heterozygous mutations in several of the more rare Fanconi anemia subtype genes—*FANCD1/BRCA2*, *FANCI/BRIP1/BACH1*, *FANCN/PALB2*, *FANCO/RAD51C*, *FANCES/BRCA1*, and *FANCU/XRCC2*—are associated with moderate adult onset cancer risks, particularly for breast and ovarian cancer. Thus, parents of children with these Fanconi anemia subtypes may benefit from increased cancer screening and prevention strategies. The use of multigene panel testing for individuals at risk for hereditary breast/ovarian cancer is increasingly identifying adult carriers who also need to be alerted to their cancer risk, as well as their risk for Fanconi anemia in their offspring and options for preconception planning and testing of their partner for the same Fanconi anemia gene (3, 69).

The Fanconi anemia family support group, Fanconi Anemia Research Fund, [www.fanconi.org](http://www.fanconi.org), has information for patients, clinicians, and researchers.

## Xeroderma Pigmentosum

### Genetic summary

Xeroderma pigmentosum (OMIMs: #278700, #278720, #278730, #278740, #278750, #610651) is caused by AR inheritance of pathogenic variants in the nucleotide excision repair genes *DDB2*, *ERCC1*, *ERCC2*, *ERCC3*, *ERCC4*, *ERCC5*, *POLH*, *XPA*, or *XPC*. Patients with xeroderma pigmentosum have severe sun sensitivity, and develop significant skin freckling and skin cancers (basal cell, skin SCC, and melanoma). Other cancers that have been described in xeroderma pigmentosum patients include leukemia, SCC (common sites face, head and neck), brain and spinal cord tumors, and other solid tumors (70–74). Eye involvement in xeroderma pigmentosum can be significant, with keratitis and lid atrophy. Some patients may have neurologic symptoms including progressive sensorineural hearing loss and cognitive impairment (70, 75–78). Xeroderma pigmentosum is a rare disorder and estimated to affect 1 in 1 million people in the United States and Europe, with a slightly increased frequency in the Middle East, Japan, and North Africa (79).

### Cancer screening/surveillance/management protocols

Screening will be most effective when paired with strategies to minimize UV exposure. Patients and families should be educated about limiting UV exposure by protecting all surfaces of the body and the eyes, and should be provided with psychosocial support to help ensure adherence to these measures. Those with xeroderma pigmentosum are most sensitive to UVA and UVB radiation, which come from the sun, but indoor light sources can also produce UV and should be evaluated with a light meter to identify sources of significant UV that could be replaced (2). Patients with xeroderma pigmentosum require multidisciplinary care including comprehensive dermatologic evaluations at least every 3 months. Close monitoring is recommended by ophthalmology for ocular disease; by otolaryngology for hearing loss; as well as endocrinology and nutrition for dietary supplementation, specifically vitamin D (2). Additional information for families with xeroderma pigmentosum is available at <http://www.xps.org/>

## References

- Gatti R, Perlman S. Ataxia-telangiectasia. In: Pagon RA, Adam MP, Ardinger HH, Wallace SE, Amemiya A, Bean LJH, et al., editors. GeneReviews

## Heterozygous Carriers of Pathogenic Variants in DNA Repair Genes

The majority of the DNA repair disorders described above are AR syndromes with a few exceptions. Heterozygous carriers of pathogenic variants in DNA repair genes may have an elevated cancer risk, but the data vary by gene and cancer. Parents of children with AR DNA repair disorders should receive genetic counseling and cancer screening in accordance with national guidelines and expert providers (24, 80).

In addition, nonsyndromic children may be heterozygous carriers for DNA damage genes, and recent next-generation sequencing studies have identified children with cancer harboring pathogenic variants in these genes (81–83). At this time, it is not clear whether these variants are directly connected to the cancer affecting the children in those studies. Providers should tailor their discussions with these families based on comprehensive patient-specific information including other clinical features of the disorder being present and other biomarker or functional evidence before determining whether testing and/or screening should be considered for unaffected siblings under the age. It should be noted, however, that genetic testing of children for adult-onset diseases is generally not recommended (84).

## Conclusions

DNA repair syndromes manifest from heritable underpinnings and when identified, affected individuals require multidisciplinary care, nuanced therapeutic considerations, and screening (Table 2). Although these syndromes are rare, dedicated syndrome registries, basic science research, and clinical research continue to develop the foundation for the most appropriate treatment and management for individuals with inherent aberrant DNA repair mechanisms. Centralized centers of excellence are highly recommended to be involved directly or through consultation in caring for patients with heritable pediatric DNA damage syndromes. For parents carrying a single mutation in a DNA damage syndrome gene, screening and prevention considerations may be indicated and necessitate genetic counseling and guidance.

## Disclosure of Potential Conflicts of Interest

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[Internet]. Seattle (WA): University of Washington, Seattle; 1993. 1999 Mar 19 [updated 2016 Oct 27].

2. Kraemer KH, DiGiovanna JJ. Xeroderma pigmentosum. In: Pagon RA, Adam MP, Ardinger HH, Wallace SE, Amemiya A, Bean LJH, et al., editors. GeneReviews [Internet]. Seattle (WA): University of Washington, Seattle; 1993. 2003 Jun 20 [updated 2016 Sep 29].
3. Mehta PA, Tolar J. Fanconi anemia. In: Pagon RA, Adam MP, Ardinger HH, Wallace SE, Amemiya A, Bean LJH, et al., editors. GeneReviews [Internet]. Seattle (WA): University of Washington, Seattle; 1993. 2002 Feb 14 [updated 2017 Feb 23].
4. Sanz MM, German J, Cunniff C. Bloom's syndrome. In: Pagon RA, Adam MP, Ardinger HH, Wallace SE, Amemiya A, Bean LJH, et al., editors. GeneReviews [Internet]. Seattle (WA): University of Washington, Seattle; 1993. 2006 Mar 22 [updated 2016 Apr 7].
5. Savage SA, Dokal I, Armanios M, Aubert G, Cowen EW, Domingo DL, et al. Dyskeratosis congenita: the first NIH clinical research workshop. *Pediatr Blood Cancer* 2009;53:520-3.
6. Varon R, Demuth I, Chrzanoswska KH. Nijmegen breakage syndrome. In: Pagon RA, Adam MP, Ardinger HH, Wallace SE, Amemiya A, Bean LJH, et al., editors. GeneReviews [Internet]. Seattle (WA): University of Washington, Seattle; 1993. 1999 May 17 [updated 2017 Feb 2].
7. Vulliamy T, Dokal I. Dyskeratosis congenita. *Semin Hematol* 2006;43:157-66.
8. Wang LL, Plon SE. Rothmund-Thomson syndrome. In: Pagon RA, Adam MP, Ardinger HH, Wallace SE, Amemiya A, Bean LJH, et al., editors. GeneReviews [Internet]. Seattle (WA): University of Washington, Seattle; 1993. 1999 Oct 6 [updated 2016 Aug 11].
9. Chun HH, Gatti RA. Ataxia-telangiectasia, an evolving phenotype. *DNA Repair* 2004;3:1187-96.
10. National Center for Biotechnology Information. ATM ATM serine/threonine. Available from: <https://www.ncbi.nlm.nih.gov/gene/472>; 2016.
11. McKinnon PJ. ATM and ataxia telangiectasia. *EMBO Rep* 2004;5:772-6.
12. Suarez F, Mahlaoui N, Canioni D, Andriamanga C, Dubois d'Enghien C, Brousse N, et al. Incidence, presentation, and prognosis of malignancies in ataxia-telangiectasia: a report from the French national registry of primary immune deficiencies. *J Clin Oncol* 2015;33:202-8.
13. Porter CC, Druley TE, Erez A, Kuiper RP, Onel K, Schiffman JD, et al. Recommendations for surveillance for children with leukemia-predisposing conditions. *Clin Cancer Res* 2017;23:e14-e22.
14. Grant RC, Selander I, Connor AA, Selvarajah S, Borgida A, Briollais L, et al. Prevalence of germline mutations in cancer predisposition genes in patients with pancreatic cancer. *Gastroenterology* 2015;148:556-64.
15. Pritchard CC, Mateo J, Walsh MF, De Sarkar N, Abida W, Beltran H, et al. Inherited DNA-repair gene mutations in men with metastatic prostate cancer. *N Engl J Med* 2016;375:443-53.
16. Walsh MF, Nathanson KL, Couch FJ, Offit K. Genomic biomarkers for breast cancer risk. *Adv Exp Med Biol* 2016;882:1-32.
17. Wegner RD, Metzger M, Hanefeld F, Jaspers NG, Baan C, Magdorf K, et al. A new chromosomal instability disorder confirmed by complementation studies. *Clin Genet* 1988;33:20-32.
18. Varon R, Vissinga C, Platzer M, Cerosaletti KM, Chrzanoswska KH, Saar K, et al. Nibrin, a novel DNA double-strand break repair protein, is mutated in Nijmegen breakage syndrome. *Cell* 1998;93:467-76.
19. Albert MH, Gennery AR, Greil J, Cale CM, Kalwak K, Kondratenko I, et al. Successful SCT for Nijmegen breakage syndrome. *Bone Marrow Transplant* 2010;45:622-6.
20. Dembowska-Baginska B, Perek D, Brozyna A, Wakulinska A, Olczak-Kowalczyk D, Gladkowska-Dura M, et al. Non-Hodgkin lymphoma (NHL) in children with Nijmegen Breakage syndrome (NBS). *Pediatr Blood Cancer* 2009;52:186-90.
21. Dumic M, Radman I, Krnic N, Nola M, Kusec R, Begovic D, et al. Successful treatment of diffuse large B-cell non-hodgkin lymphoma with modified CHOP (cyclophosphamide/doxorubicin/vincristine/prednisone) chemotherapy and rituximab in a patient with Nijmegen syndrome. *Clin Lymphoma Myeloma* 2007;7:590-3.
22. Jonkman-Berk BM, van den Berg JM, Ten Berge IJ, Bredius RG, Driessen GJ, Dalm VA, et al. Primary immunodeficiencies in the Netherlands: national patient data demonstrate the increased risk of malignancy. *Clin Immunol* 2015;156:154-62.
23. Daly MB, Pilarski R, Berry M, Buys SS, Farmer M, Friedman S, et al. NCCN Guidelines insights: genetic/familial high-risk assessment: breast and ovarian version 2.2015. *J Natl Compr Canc Netw* 2017;15:9-20.
24. Bachrati CZ, Hickson ID. RecQ helicases: suppressors of tumorigenesis and premature aging. *Biochem J* 2003;374:577-606.
25. Pedrazzi G, Bachrati CZ, Selak N, Studer I, Petkovic M, Hickson ID, et al. The Bloom's syndrome helicase interacts directly with the human DNA mismatch repair protein hMSH6. *Biol Chem* 2003;384:1155-64.
26. Bloom D. Congenital telangiectatic erythema resembling lupus erythematosus in dwarfs; probably a syndrome entity. *AMA Am J Dis Child* 1954;88:754-8.
27. German J, Archibald R, Bloom D. Chromosomal breakage in a rare and probably genetically determined syndrome of man. *Science* 1965;148:506-7.
28. German J, Sanz MM, Ciocci S, Ye TZ, Ellis NA. Syndrome-causing mutations of the BLM gene in persons in the Bloom's Syndrome Registry. *Hum Mutat* 2007;28:743-53.
29. Cunniff C, Bassetti JA, Ellis NA. Bloom's syndrome: clinical spectrum, molecular pathogenesis, and cancer predisposition. *Mol Syndromol* 2017;8:4-23.
30. Kitao S, Lindor NM, Shiratori M, Furuichi Y, Shimamoto A. Rothmund-thomson syndrome responsible gene, RECQL4: genomic structure and products. *Genomics* 1999;61:268-76.
31. Kitao S, Shimamoto A, Goto M, Miller RW, Smithson WA, Lindor NM, et al. Mutations in RECQL4 cause a subset of cases of Rothmund-Thomson syndrome. *Nat Genet* 1999;22:82-4.
32. National Center for Biotechnology Information. RECQL4 RecQ like helicase 4 [Homo sapiens (human)]. Available from: <https://www.ncbi.nlm.nih.gov/gene?Db=gene&Cmd=DetailsSearch&Term=641>.
33. Lu L, Jin W, Wang LL. Aging in Rothmund-Thomson syndrome and related RECQL4 genetic disorders. *Ageing Res Rev* 2017;33:30-5.
34. Moss C. Rothmund-Thomson syndrome: a report of two patients and a review of the literature. *Br J Dermatol* 1990;122:821-9.
35. Moss C. "New" syndrome with telangiectasia, dwarfism, and spondyloepiphyseal dysplasia may be Rothmund-Thomson syndrome. *Pediatr Dermatol* 1990;7:82-4.
36. Wang LL, Levy ML, Lewis RA, Chintagumpala MM, Lev D, Rogers M, et al. Clinical manifestations in a cohort of 41 Rothmund-Thomson syndrome patients. *Am J Med Genet* 2001;102:11-7.
37. Haytac MC, Oztunc H, Mete UO, Kaya M. Rothmund-Thomson syndrome: a case report. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2002;94:479-84.
38. Mehollin-Ray AR, Kozinetz CA, Schlesinger AE, Guilleman RP, Wang LL. Radiographic abnormalities in Rothmund-Thomson syndrome and genotype-phenotype correlation with RECQL4 mutation status. *AJR Am J Roentgenol* 2008;191:W62-6.
39. Hicks MJ, Roth JR, Kozinetz CA, Wang LL. Clinicopathologic features of osteosarcoma in patients with Rothmund-Thomson syndrome. *J Clin Oncol* 2007;25:370-5.
40. Wang LL, Gannavarapu A, Kozinetz CA, Levy ML, Lewis RA, Chintagumpala MM, et al. Association between osteosarcoma and deleterious mutations in the RECQL4 gene in Rothmund-Thomson syndrome. *J Natl Cancer Inst* 2003;95:669-74.
41. Simon T, Kohlhase J, Wilhelm C, Kochanek M, De Carolis B, Berthold F. Multiple malignant diseases in a patient with Rothmund-Thomson syndrome with RECQL4 mutations: case report and literature review. *Am J Med Genet A* 2010;152A:1575-9.
42. Siitonen HA, Sotkasiira J, Biervliet M, Benmansour A, Capri Y, Cormier-Daire V, et al. The mutation spectrum in RECQL4 diseases. *Eur J Hum Genet* 2009;17:151-8.
43. Knoell KA, Sidhu-Malik NK, Malik RK. Aplastic anemia in a patient with Rothmund-Thomson syndrome. *J Pediatr Hematol Oncol* 1999;21:444-6.
44. Narayan S, Fleming C, Trainer AH, Craig JA. Rothmund-Thomson syndrome with myelodysplasia. *Pediatr Dermatol* 2001;18:210-2.
45. Pianigiani E, De Aloe G, Andreassi A, Rubegni P, Fimiani M. Rothmund-Thomson syndrome (Thomson-type) and myelodysplasia. *Pediatr Dermatol* 2001;18:422-5.
46. Porter WM, Hardman CM, Abdalla SH, Powles AV. Haematological disease in siblings with Rothmund-Thomson syndrome. *Clin Exp Dermatol* 1999;24:452-4.
47. Spurney C, Gorlick R, Meyers PA, Healey JH, Huvos AG. Multicentric osteosarcoma, Rothmund-Thomson syndrome, and secondary



- nasopharyngeal non-Hodgkin's lymphoma: a case report and review of the literature. *J Pediatr Hematol Oncol* 1998;20:494-7.
48. Shuttleworth D, Marks R, Griffin PJ, Salaman JR. Treatment of cutaneous neoplasia with etretinate in renal transplant recipients. *Q J Med* 1988;68:717-25.
  49. Potozkin JR, Geronemus RG. Treatment of the poikilodermatous component of the Rothmund-Thomson syndrome with the flashlamp-pumped pulsed dye laser: a case report. *Pediatr Dermatol* 1991;8:162-5.
  50. Bertuch AA. The molecular genetics of the telomere biology disorders. *RNA Biol* 2016;13:696-706.
  51. Alter BP, Giri N, Savage SA, Peters JA, Loud JT, Leathwood L, et al. Malignancies and survival patterns in the National Cancer Institute inherited bone marrow failure syndromes cohort study. *Br J Haematol* 2010;150:179-88.
  52. Alder JK, Chen JJ, Lancaster L, Danoff S, Su SC, Cogan JD, et al. Short telomeres are a risk factor for idiopathic pulmonary fibrosis. *Proc Natl Acad Sci U S A* 2008;105:13051-6.
  53. Armanios MY, Chen JJ, Cogan JD, Alder JK, Ingersoll RG, Markin C, et al. Telomerase mutations in families with idiopathic pulmonary fibrosis. *N Engl J Med* 2007;356:1317-26.
  54. de la Fuente J, Dokal I. Dyskeratosis congenita: advances in the understanding of the telomerase defect and the role of stem cell transplantation. *Pediatr Transplant* 2007;11:584-94.
  55. Ameziane N, May P, Haitjema A, van de Vrugt HJ, van Rossum-Fikkert SE, Ristic D, et al. A novel Fanconi anaemia subtype associated with a dominant-negative mutation in RAD51. *Nat Commun* 2015;6:8829.
  56. Brosh RM Jr, Bellani M, Liu Y, Seidman MM. Fanconi anemia: a DNA repair disorder characterized by accelerated decline of the hematopoietic stem cell compartment and other features of aging. *Ageing Res Rev* 2017;33:67-75.
  57. Faivre L, Guardiola P, Lewis C, Dokal I, Ebell W, Zatterale A, et al. Association of complementation group and mutation type with clinical outcome in fanconi anemia. European Fanconi Anemia Research Group. *Blood* 2000;96:4064-70.
  58. Solomon PJ, Margaret P, Rajendran R, Ramalingam R, Menezes GA, Shirley AS, et al. A case report and literature review of Fanconi anemia (FA) diagnosed by genetic testing. *Ital J Pediatr* 2015;41:38.
  59. Andreassen PR, D'Andrea AD, Taniguchi T. ATR couples FANCD2 monoubiquitination to the DNA-damage response. *Genes Dev* 2004;18:1958-63.
  60. Kottmann MC, Smogorzewska A. Fanconi anaemia and the repair of Watson and Crick DNA crosslinks. *Nature* 2013;493:356-63.
  61. Auerbach AD. Fanconi anemia and leukemia: tracking the genes. *Leukemia* 1992;6Suppl 1:1-4.
  62. Auerbach AD, Rogatko A, Schroeder-Kurth TM. International Fanconi Anemia Registry: relation of clinical symptoms to diepoxybutane sensitivity. *Blood* 1989;73:391-6.
  63. Giampietro PF, Adler-Brecher B, Verlander PC, Pavlakis SG, Davis JG, Auerbach AD. The need for more accurate and timely diagnosis in Fanconi anemia: a report from the International Fanconi Anemia Registry. *Pediatrics* 1993;91:1116-20.
  64. Alter BP. Fanconi anemia and the development of leukemia. *Best Pract Res Clin Haematol* 2014;27:214-21.
  65. Alter BP, Olson SB. Wilms tumor, AML, and medulloblastoma in a child with cancer prone syndrome of total premature chromatid separation and Fanconi anemia. *Pediatr Blood Cancer* 2010;54:488.
  66. Kutler DI, Auerbach AD, Satagopan J, Giampietro PF, Batish SD, Huvos AG, et al. High incidence of head and neck squamous cell carcinoma in patients with Fanconi anemia. *Arch Otolaryngol Head Neck Surg* 2003;129:106-12.
  67. Kutler DI, Singh B, Satagopan J, Batish SD, Berwick M, Giampietro PF, et al. A 20-year perspective on the International Fanconi Anemia Registry (IFAR). *Blood* 2003;101:1249-56.
  68. Rosenberg PS, Greene MH, Alter BP. Cancer incidence in persons with Fanconi anemia. *Blood* 2003;101:822-6.
  69. National Comprehensive Cancer Network. HBOC Guidelines 2016. Available from: [https://www.nccn.org/professionals/physician\\_gls/f\\_guidelines.asp](https://www.nccn.org/professionals/physician_gls/f_guidelines.asp).
  70. Kraemer KH, Lee MM, Scotto J. Xeroderma pigmentosum. Cutaneous, ocular, and neurologic abnormalities in 830 published cases. *Arch Dermatol* 1987;123:241-50.
  71. Bradford PT, Goldstein AM, Tamura D, Khan SG, Ueda T, Boyle J, et al. Cancer and neurologic degeneration in xeroderma pigmentosum: long term follow-up characterises the role of DNA repair. *J Med Genet* 2011;48:168-76.
  72. Butt FM, Moshi JR, Owibingire S, Chindia ML. Xeroderma pigmentosum: a review and case series. *J Craniomaxillofac Surg* 2010;38:534-7.
  73. DiGiovanna JJ, Patronas N, Katz D, Abangan D, Kraemer KH. Xeroderma pigmentosum: spinal cord astrocytoma with 9-year survival after radiation and isotretinoin therapy. *J Cutan Med Surg* 1998;2:153-8.
  74. Fassih H, Sethi M, Fawcett H, Wing J, Chandler N, Mohammed S, et al. Deep phenotyping of 89 xeroderma pigmentosum patients reveals unexpected heterogeneity dependent on the precise molecular defect. *Proc Natl Acad Sci U S A* 2016;113:E1236-45.
  75. Kraemer KH, Lee MM, Andrews AD, Lambert WC. The role of sunlight and DNA repair in melanoma and nonmelanoma skin cancer. The xeroderma pigmentosum paradigm. *Arch Dermatol* 1994;130:1018-21.
  76. Kraemer KH, Levy DD, Parris CN, Gozukara EM, Moriwaki S, Adelberg S, et al. Xeroderma pigmentosum and related disorders: examining the linkage between defective DNA repair and cancer. *J Invest Dermatol* 1994;103(5 Suppl):96S-101S.
  77. Rapin I, Lindenbaum Y, Dickson DW, Kraemer KH, Robbins JH. Cockayne syndrome and xeroderma pigmentosum. *Neurology* 2000;55:1442-9.
  78. Totonchy MB, Tamura D, Pantell MS, Zalewski C, Bradford PT, Merchant SN, et al. Auditory analysis of xeroderma pigmentosum 1971-2012: hearing function, sun sensitivity and DNA repair predict neurological degeneration. *Brain* 2013;136:194-208.
  79. Kleijer WJ, Laugel V, Berneburg M, Nardo T, Fawcett H, Gratchev A, et al. Incidence of DNA repair deficiency disorders in western Europe: Xeroderma pigmentosum, Cockayne syndrome and trichothiodystrophy. *DNA Repair* 2008;7:744-50.
  80. Tung N, Domchek SM, Stadler Z, Nathanson KL, Couch F, Garber JE, et al. Counselling framework for moderate-penetrance cancer-susceptibility mutations. *Nat Rev Clin Oncol* 2016;13:581-8.
  81. Mody RJ, Wu YM, Lonigro RJ, Cao X, Roychowdhury S, Vats P, et al. Integrative clinical sequencing in the management of refractory or relapsed cancer in youth. *JAMA* 2015;314:913-25.
  82. Parsons DW, Roy A, Yang Y, Wang T, Scollon S, Bergstrom K, et al. Diagnostic yield of clinical tumor and germline whole-exome sequencing for children with solid tumors. *JAMA Oncol* 2016 Jan 28. [Epub ahead of print].
  83. Zhang J, Walsh MF, Wu G, Edmonson MN, Gruber TA, Easton J, et al. Germline mutations in predisposition genes in pediatric cancer. *N Engl J Med* 2015;373:2336-46.
  84. Kalia SS, Adelman K, Bale SJ, Chung WK, Eng C, Evans JP, et al. Recommendations for reporting of secondary findings in clinical exome and genome sequencing, 2016 update (ACMG SF v2.0): a policy statement of the American College of Medical Genetics and Genomics. *Genet Med* 2017;19:249-55.

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## Recommendations for Childhood Cancer Screening and Surveillance in DNA Repair Disorders

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