Cancer Screening Recommendations for Individuals with Li-Fraumeni Syndrome

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Abstract

Li-Fraumeni syndrome (LFS) is an autosomal dominantly inherited condition caused by germline mutations of the *TP53* tumor suppressor gene encoding p53, a transcription factor triggered as a protective cellular mechanism against different stressors. Loss of p53 function renders affected individuals highly susceptible to a broad range of solid and hematologic cancers. It has recently become evident that children and adults with LFS benefit from intensive surveillance aimed at early tumor detection. In October 2016, the American Association for Cancer Research held a meeting of international LFS experts to evaluate the current knowledge on LFS and propose consensus surveillance recommendations. Herein, we briefly summarize clinical and genetic aspects of this aggressive cancer predisposition syndrome. In addition, the expert panel concludes that there are sufficient existing data to recommend that all patients with LFS be offered cancer surveillance as soon as the clinical or molecular LFS diagnosis is established. Specifically, the panel recommends adoption of a modified version of the "Toronto protocol" that includes a combination of physical exams, blood tests, and imaging. The panel also recommends that further research be promoted to explore the feasibility and effectiveness of these risk-adapted surveillance and cancer prevention strategies while addressing the psychosocial needs of individuals and families with LFS. *Clin Cancer Res; 23(11); e38–e45. ©2017 AACR.*

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Introduction

Li-Fraumeni syndrome (LFS; OMIM #151623) is among the most aggressive cancer predisposition syndromes characterized by a high and early-onset cancer risk. The tumor spectrum is wide and includes brain tumors [choroid plexus carcinoma, Sonic Hedgehog (SHH) subtype medulloblastoma, glioma], adrenocortical carcinoma (ACC), a range of soft tissue sarcomas (STS) and bone tumors, hematologic malignancies, breast cancer (generally very early in onset), and other cancer types, including lung, skin, gastrointestinal tract, kidney, thyroid, as well as neuroblastoma. The tumors most closely associated with LFS are called "core" cancers and include STS, osteosarcoma, premenopausal breast cancer, brain tumors, and ACCs (for review, see refs. 1, 2).

LFS was first described in 1969 by Frederick Li and Joseph Fraumeni Jr based on their observation of a unique spectrum of cancers in four families in whom the index cases presented with rhabdomyosarcoma (3). The original definition of the syndrome was established in 1988 as the result of an analysis of 24 kindreds presenting with an autosomal dominant pattern of transmission of early-onset neoplasms including STS, breast cancers, central nervous system (CNS) tumors, leukemias, and ACCs before the age of 45 years (4). This "classical" definition requires one individual with a sarcoma diagnosed under the age of 45 who has at least one first-degree relative (parent, sibling, or child) with a cancer of any kind diagnosed under the age of 45 and a third family member who is either a first- or second-degree relative in the same parental lineage (grandparent, aunt, uncle, niece, nephew, or grandchild) with any cancer diagnosed under the age of 45, or a sarcoma at any age (4).

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In 1990, germline *TP53* mutations were discovered as the only cause of LFS (5–7). The identification of germline *TP53* mutations in patients not fulfilling the original definition of the syndrome led to periodic updates of operational LFS criteria, designated the "Chompret criteria," to describe four different clinical situations with a high probability of being caused by an underlying *TP53* mutation and in which genetic counseling and clinical *TP53* mutation testing should be strongly considered and offered: (i) familial presentation: proband with an LFS spectrum tumor (premenopausal breast cancer, STS, brain tumor, ACC) prior to age 46 years and at least one first- or second-degree relative with an LFS tumor (except breast cancer, if the proband has breast cancer) before the age of 56 years or with multiple tumors; (ii) multiple tumors: proband with multiple malignancies (except two breast cancers), of which at least two belong to the LFS spectrum, before the age of 46 years; (iii) rare tumors: patients with ACC, choroid plexus carcinoma, or embryonal anaplastic subtype rhabdomyosarcoma independent of family history; and (iv) breast cancer before the age of 31 years (8–12).

Recent sequencing projects have shown that LFS plays a significant role in the pathogenesis of childhood cancers. Eighty percent of children with rhabdomyosarcoma with diffuse anaplasia (12), 50% of children with ACC (13), 40% of children with choroid plexus carcinoma (11), 40% of children with low-hypodiploid acute lymphoblastic leukemia (ALL; ref. 14), more than 10% of children with SHH medulloblastoma (15, 16), up to 10% of children with osteosarcoma (17–19), and 1% to 2% of children with relapsed ALL have a germline *TP53* mutation, often in the absence of an obvious family history (20). Also, rare *TP53* germline variants contribute to the development of childhood neuroblastoma (21). We expect that future research projects analyzing the germline DNA sequence of children with cancer will reveal a more complete childhood cancer spectrum of LFS. Multigene panel germline testing of adults with cancer or a positive family cancer history may also expand the phenotypic picture of LFS (22). Of note, *TP53* germline mutations lead to specific somatic aberrations and mutation signatures in LFS-related cancers (23). Consequently, detection of such signatures, such as excessive chromothripsis in medulloblastoma, should raise the suspicion of an underlying *TP53* germline mutation (24).

Penetrance

Ascertainment bias is likely to lead to an overestimation of tumor risk in individuals with LFS, and future studies will be required to provide more accurate cancer risk estimates. It is important to note that individuals with a germline *TP53* mutation display great clinical heterogeneity in terms of cancer type and age of onset (25–27). A recent study described 214 LFS families diagnosed between 1993 and 2013 and included 415 constitutional *TP53* mutation carriers (11, 28), 322 (78%) of whom developed at least one malignancy. A significant number of cancers occurred at a young age; namely, 22% were diagnosed with a cancer by age 5 years and 41% by age 18 years (11). Notably, 4% of participants developed a malignancy during the first year of life (11). In children and adolescents with LFS, osteosarcoma was the most common tumor (30%), followed by ACC (27%), brain tumors (25%), and STS (23%; ref. 11). Breast cancer was the most frequently encountered malignancy (79% of women), followed by STS (27%) in adults with LFS. Second neoplasms occurred in 40% of patients, often within the radiation field, which is in agreement with previous observations and with the notion that initial antitumor therapy increases the risk of subsequent cancers (29–34).

Investigators from the NCI (Bethesda, Maryland) recently evaluated 286 *TP53* mutation–positive individuals from 107 families (35). The cumulative cancer incidence was 50% by age 31 years among females with a *TP53* mutation and 50% by 46 years among males, and nearly 100% by age 70 years for the entire cohort (35). Cancer risk was highest after age 20 years for females, mostly due to breast cancer. Among males, the risk was higher in childhood and later adulthood (35). Among females, the cumulative incidence rates by age 70 years were 54% for breast cancer, 15% for STS, 6% for brain tumors, and 5% for osteosarcoma. Among males, the incidence rates were 22% for STS, 19% for brain tumors, and 11% for osteosarcoma (35). After a median of 10 years, almost 50% of those with one cancer developed at least one other cancer (35). As noted above, these estimates also likely suffer from an ascertainment bias, as most of the *TP53* analyses have been performed in affected children with familial history of cancer or multiple primaries. With the exponential increase of *TP53* tests being performed in cancer patients, germline *TP53* mutations are now more frequently identified in patients and families who have developed only adult cancers (22). Notably, however, a pattern of genetic anticipation is frequently observed in individual LFS families. The underlying genetic mechanisms remain unknown (36).

Brazilian founder mutation

In Brazil, a high prevalence of LFS is present due to a founder effect mutation. A germline *TP53* mutation (c.1010G>A; p.R337H) is present in 0.3% of individuals from the South/Southeastern regions, and it is estimated that more than 300,000 Brazilian individuals have LFS. The spectrum of cancers occurring in carriers is similar to the cancer spectrum observed in patients who carry other *TP53* mutations and includes STS, early-onset breast cancer, cancers of the CNS, and childhood ACC. However, p.R337H carriers have a higher occurrence of young adult papillary thyroid cancer, renal cancer, and lung adenocarcinoma than carriers of other *TP53* mutations (37). Assessment of pedigrees and familial cancer patterns shows significant differences between p.R337H and classic *TP53* mutation carriers. The penetrance of cancer before age 30 is estimated to be 15% to 20% compared with 50% in carriers of classic mutations (38). Also, tumor patterns are different from those documented in other *TP53* mutation carriers. ACCs represent over 8% of all tumors in p.R337H carriers (compared with 4% for classic mutations). Furthermore, adult tumor onset is later in p.R337H carriers. Breast cancers occur at a mean age of 40 years—later than in classic carriers in whom the mean age of onset is 32 years. Although the familial presentation of cancer risk in p.R337H mutation carriers is within the LFS spectrum, the occurrence of specific traits that are unique to the carriers of the Brazilian founder mutation may suggest it represents a variant form of LFS (39, 40).

Genetic Summary

TP53 function and phenotype–genotype correlation

The *TP53* gene encodes a transcription factor that is activated in response to a variety of cellular stress factors and controls the expression of multiple genes that govern cellular processes crucial for tumor suppression (41). More than 250 different *TP53* germline alterations have been reported, and the types of mutations

resemble those that occur as somatic events (13, 42). Missense mutations are the most common, occurring in approximately 70% of cases and most often altering residues within the DNA-binding domain (13, 42). Other alterations and defects exist (splicing, intragenic deletion, frameshift, nonsense, in-frame insertion/deletion, intronic; ref. 13). Twenty percent of LFS families harbor one of six hotspot mutations (p.R175H, p.G245S, p.R248Q, p.R248W, p.R273H, and p.R282W; ref. 13), and the rate of *de novo* mutations could be as high as 25% (43). The *TP53* germline mutation type and its effect on p53 function may influence disease penetrance as well as the cancer site and the risk of secondary malignancies. The highest cancer risk is associated with dominant-negative *TP53* missense mutations within the DNA-binding domain. Such mutations are detected commonly in LFS patients with brain tumors (62%), osteosarcoma (40%), and rhabdomyosarcoma (36%; ref. 11). Non–dominant-negative *TP53* mutations occur more frequently in patients with ACC (76%; refs. 11, 28). Not only specific mutations but also genetic modifiers are proposed to influence the LFS phenotype. These modifiers include the *MDM2* polymorphism rs2279744 (44); *TP53* polymorphisms, such as a duplication within intron 3 (PIN3; refs. 45, 46); telomere length (47); differential methylation or variant alleles in miRNAs that modify p53-mediated cell regulation (48–50); and the accumulation of copy number variations (CNV; ref. 51).

Cancer Screening/Surveillance Protocols

In recent years, with the aim of early tumor detection and reduction of cancer and treatment-related morbidity and mortality, suggestions for clinical surveillance of *TP53* mutation carriers have been proposed from Australia, the United States [National Comprehensive Cancer Network (NCCN) Guidelines], and Canada (Table 1; refs. 52–56). Over an 11-year period, investigators in Toronto, Salt Lake City, and Los Angeles (subsequently Columbus) prospectively followed and reported on the feasibility and outcomes of screening children and adults using a multi-modality protocol that has been coined the "Toronto protocol" (Table 1; refs. 55, 56). In patients who decided to undergo surveillance, compliance with key components of the protocol, including whole-body MRI (WBMRI) and brain MRI, was shown to be >90%. Forty tumors were detected in 19 of 59 patients on surveillance over a median follow-up period of 32 months in contrast to 61 tumors that presented in 43 of 49 patients who did not undergo surveillance. Furthermore, 25 of 40 tumors on the surveillance "arm" were low grade or premalignant at the time of detection, suggesting that early detection through surveillance may identify lesions before malignant transformation. Importantly, an improved overall survival (OS) was observed in individuals undergoing surveillance: 5-year OS, 88.8% versus 59.6% (surveillance vs. nonsurveillance groups). Different screening modalities identified the spectrum of tumors, and although WBMRI (as distinct from dedicated brain or breast MRI) was an important component, only 20% of tumors (e.g., malignant fibrous histiocytoma, osteosarcoma, osteochondroma, ACC, chordoma, chondroma, colorectal carcinoma, and lung carcinoma) were detected by that modality. In addition, WBMRI identified two biopsy-proven nontumor lesions (false positives), and two false negative scans were reported (56). Thus, although WBMRI comprises a critical component of LFS surveillance, it is important to keep in mind that it is meant as a screening tool to be

employed in conjunction with the other components of the Toronto protocol [see article by Greer and colleagues in this series (57)]. The outcome of other screening modalities included two ACCs detected with abdominal ultrasound (US) and ACC-specific blood work. A third ACC was detected with WBMRI. All were diagnosed in children <10 years of age. Eight CNS lesions were detected with brain MRI: choroid plexus carcinoma (two), diffuse astrocytoma (two), low-grade glioma (one), meningioma (one), dysembryoplastic neuroepithelial tumor (one), and subependymoma (one). Clinical exam contributed to the diagnosis of eight tumors. Colonoscopy detected five precancerous adenomas in four individuals and one rectal adenocarcinoma. However, outside of ACC screening early in life, abdominal and pelvic US did not detect any cancers (56).

[^{18}F] 2-fluoro-2deoxy-D-glucose (^{18}F-FDG) PET/CT imaging (F^{18}-FDG PET/CT) as a screening tool has been employed by two groups (58, 59) and was able to detect asymptomatic malignancies. However, because this technology necessitates radiation exposure, there is less enthusiasm for its use as a cancer screening tool. In a Brazilian study, neonatal screening for the p.R337H mutation followed by surveillance in mutation carriers (medical exam, dehydroepiandrosterone/total testosterone/cortisol assay, abdominal US; frequency determined by age) led to the detection of ACCs diagnosed at lower stages (seven ACCs, all stage I) but failed to offer adequate screening to parents and adult family members harboring the same mutation (60).

Key issues related to psychosocial and other impacts on children undergoing surveillance and parents/family members who may be at risk

There is a paucity of data on the psychosocial impact (to both affected individuals and relatives) of LFS testing and a clear need for future research in this field, performed in conjunction with prospective surveillance programs (61–69). Traditionally, LFS families with a history of multiple malignancies have been felt to carry a significant psychological burden given their exposure to multiple experiences of grief and threats to personal well-being (70). Many families undergoing surveillance believe in the value of this approach to detect tumors at an early stage or grade, reporting enhanced sense of control, security, and empowerment (63, 71). However, intense cancer surveillance schedules may be a burden to other families: Not all LFS patients choose to participate in the screening, and some find it too anxiety provoking. As is the case with the management of patients with any cancer predisposition syndrome, screening and surveillance strategies impose physical, psychosocial, and financial challenges to patients and families. With the wide adoption of next-generation sequencing (NGS) panels, many individuals with *TP53* mutations lack classic personal or family history of LFS-related cancers. This may create a different set of psychosocial issues. As described in more detail by Druker and colleagues (72) in this *CCR* Pediatric Oncology Series, these challenges are best explored and managed in the context of multidisciplinary care teams, including physicians, nurses, psychologists, and genetic counselors, with the added support of family/patient advocates and stakeholder communities.

Summary of surveillance studies recommended by the authors
Who should have surveillance? Data strongly indicate that surveillance leads to early detection of cancer and significantly improves OS (55, 56). Therefore, the expert panel recommends that surveillance should be offered to the following: (i) individuals

Table 1. Published surveillance protocols for individuals with LFS

Tumor type	Australia (52, 53)	NCCN (54)	Toronto (55, 56)
ACC	AUS q 3–4 m: birth–10 y	No screening described	• AUS q 3-4 m: birth–40 y • Biochemistry (17-OH-progesterone, total testosterone, DHEAS, androstenedione) q 3–4 m: birth–40 y • 24-h urine cortisol, if feasible
Breast cancer	• BSE: from 18 y • CBE q 6–12 m: from 20–25 y • Breast MRI annually: 20/25–50 y • (Consider annual mammography ± US if not possible) • Discuss risk-reducing bilateral mastectomy	• Breast awareness: from 18 y • CBE q 6–12 m: from 20–25 y • 20–29 y: breast MRI with contrast annually (or mammogram if unavailable) • 30–75 y: breast MRI with contrast and mammogram annually • 75 y: individual recommendations • Continue screening breast cancer survivors with mammogram and breast MRI • Discuss risk-reducing mastectomy	• BSE monthly: from 18 y • CBE q 6 m: from 20–25 y or 5–10 y before earliest case of breast cancer in family • Annual mammography and breast MRI: from age 20–75 y or 5–10 y before earliest case of breast cancer in family • Breast MRI alternates with WBMRI • Breast US with mammography as indicated by breast density • Consider risk-reducing bilateral mastectomy
Brain tumor	• Brain MRI included in annual WBMRI: potentially from childhood • Annual neurologic exam • Prompt reporting of new neurologic symptoms	• The brain may be examined as part of WBMRI or as a separate exam	• Annual brain MRI: from birth
Sarcoma	• Annual WBMRI • Annual comprehensive physical exam • Awareness of new symptoms	• Annual WBMRI (or equivalent)	• Annual rapid WBMRI: from birth • AUS q 3-4 m: from 18 y
Hematopoietic	• Annual CBC: from 18 y	• No screening described	• CBC, ESR, LDH q3-4m: from birth
CRC	• Colonoscopy q 2–5 y: from age 25 or 10 y before earliest onset of CRC in family	• Consider colonoscopy q 2–5 y: from age 25 or 5 y before earliest known colon cancer in family	• Colonoscopy q 2 y: from age 25 or 10 y before earliest onset of CRC in family
Gastric cancer	• Endoscopy q 2–5 y: from age 25 or 10 y before earliest onset gastric cancer in family		No screening described
Skin cancer	No screening described	• Annual dermatologic exam	• Annual dermatologic exam: from 18 y
Other		• Annual comprehensive physical exam, including neurologic exam • Education regarding signs and symptoms of cancer. Apprise pediatricians of childhood cancer risk • Additional surveillance based on family history of cancer • Therapeutic RT should be avoided when possible	• Complete physical exam q 3–4 m, including comprehensive neurologic exam and anthropometric measurements in children • Prompt assessment with primary care physician for any medical concerns

Abbreviations: AUS, abdominal US (abdomen and pelvis); BSE, breast self-examination; CBC, complete blood count; CBE, clinical breast examination; CRC, colorectal carcinoma; DHEAS, dehydroepiandrosterone; ESR, erythrocyte sedimentation rate; h, hour; LDH, lactate dehydrogenase; m, months; q, every; RT, radiation therapy; y, years.

carrying a pathogenic *TP53* variant and (ii) individuals fitting the "classic clinical definition" of LFS, without a pathogenic *TP53* variant.

What tests and how often? The expert panel emphasizes the central importance of a targeted history and regular physical examination in the context of potential manifestations of LFS (including glucocorticoid and sex steroid excess and neurologic changes). The expert panel recommends the use of the Toronto protocol with modifications (Table 2) for all patients, recognizing that more reliable phenotype–genotype data may lead to genotype-specific modifications of these recommendations in the future (73). Given the high ACC risk in children with LFS, we recommend US of abdomen and pelvis every 3 to 4 months until age 18 years. ACC-specific blood tests every 3 to 4 months (total testosterone, dehydroepiandrosterone sulfate, and androstenedione)

are recommended in case of a technically unsatisfactory US only. The authors suggest omission of specific ACC surveillance in adults, given its low incidence in this age group. The lifelong brain tumor risk justifies annual brain MRI. If the initial MRI performed with a gadolinium-based contrast agent (GBCA) shows normal results, the following MRIs may be conducted without GBCA unless an abnormality is seen. This is to minimize the potential for gadolinium accumulation in the basal ganglia in individuals undergoing multiple enhanced MRIs (74–76). Because of the sarcoma risk, we recommend lifelong annual WBMRI, including limbs (head to toe) and abdominal and pelvic US (every 3–4 months in children and annually in adults; every 6 months WBMRI or US). Annual WBMRI may alternate with annual dedicated brain MRI (every year, two MRIs total). However, in infants and children requiring anesthesia, and to minimize the number of health care visits, performing both MRI

Table 2. Recommended LFS screening protocol [based on the Toronto Protocol (55, 56)]

Children (birth to age 18 years)

General assessment
- Complete physical examination every 3–4 months, including blood pressure, anthropometric measurements plotted on a growth curve (with particular attention to rapid acceleration in weight or height), Cushingoid appearance, signs of virilization (pubic hair, axillary moisture, adult body odor, androgenic hair loss, clitoromegaly, or penile growth), and full neurologic assessment
- Prompt assessment with primary care physician for any medical concerns

ACC
- US of abdomen and pelvis every 3–4 months
- In case of unsatisfactory US, blood tests[a,b] may be performed every 3–4 months: total testosterone, dehydroepiandrosterone sulfate, and androstenedione

Brain tumor
- Annual brain MRI (first MRI with contrast; thereafter without contrast if previous MRI normal and no new abnormality)

Soft tissue and bone sarcoma
- Annual WBMRI

Adults

General assessment
- Complete physical examination every 6 months
- Prompt assessment with primary care physician for any medical concerns

Breast cancer
- Breast awareness (age 18 years onward)
- Clinical breast examination twice a year (age 20 years onward)
- Annual breast MRI screening[c] (ages 20–75)
- Consider risk-reducing bilateral mastectomy

Brain tumor (age 18 years onward)
- Annual brain MRI (first MRI with contrast; thereafter without contrast if previous MRI normal)

Soft tissue and bone sarcoma (age 18 years onward)
- Annual WBMRI[c]
- US of abdomen and pelvis every 12 months

Gastrointestinal cancer (age 25 years onward)
- Upper endoscopy and colonoscopy every 2–5 years

Melanoma (age 18 years onward)
- Annual dermatologic examination

Abbreviation: WBMRI, whole-body MRI, head to toe, including entire upper and lower extremities.
[a]Serial specimens obtained at the same time of day and processed in the same laboratory.
[b]The efficacy of biochemical surveillance for detection of adrenocortical carcinoma has not been shown.
[c]Breast MRI/US of abdomen and pelvis to alternate with annual WBMRI (at least one scan every 6 months).

studies concurrently every 12 months may be more appropriate. WBMRI parameters are discussed in more detail in the article by Greer and colleagues (57) in this series. Although the focus of the expert panel was on surveillance strategies for childhood-onset cancers in the context of LFS, there was general agreement that consideration of the lifelong cancer risk cannot be completely dissociated from this discussion. In light of the high early-onset breast cancer risk, we propose breast awareness (age 18 years onward) and clinical breast examination twice a year starting at age 20 years onward. In addition, from age 20 to 75 years, annual breast MRI screening is recommended, ideally alternating with annual WBMRI (one scan every 6 months). The recommendations for the use of breast US and mammography have been omitted. Also, the option of risk-reducing bilateral mastectomy should be considered and discussed with female patients. For those families

in which there was already a case of breast cancer at or around 20, awareness and screening can be considered to begin 5 to 10 years before the earliest age of onset. Because of gastrointestinal cancer risk, we propose upper endoscopy and colonoscopies every 2 to 5 years (starting at age 25 years or 5 years before the earliest age of onset in the family). Given an increased melanoma risk, annual dermatologic examinations are recommended starting at 18 years of age. Blood work for hematopoietic malignancies [namely complete blood count (CBC), erythrocyte sedimentation rate (ESR), and lactate dehydrogenase (LDH)] can be omitted because of the lack of data suggesting that presymptomatic diagnosis of leukemia leads to improved survival [see article by Porter and colleagues in this *CCR* Pediatric Oncology Series (77)]. However, for patients who received leukemogenic agents for treatment of their first cancer, consideration may be given to ongoing periodic CBCs for detection of evidence for accelerated myelodysplasia as a precursor for leukemic transformation.

When to start, when to stop? With the knowledge accumulated so far, cancer risk in children is still difficult to evaluate. As a measure of precaution, while waiting for more definitive data, the expert panel strongly advocates for proposed lifelong screening, starting as soon as a genetic diagnosis (proven *TP53* mutation carrier status) or clinical diagnosis (phenotype fits classic LFS definition) has been established. Where feasible, screening should also continue following diagnosis of a primary malignancy and be integrated with clinically indicated cancer-specific follow-up. In families with a known *TP53* germline mutation, presymptomatic testing may be offered soon after birth to begin screening within the first months of life.

Should surveillance change over time? The cancer spectrum is, at least in part, age dependent. As per the modified Toronto protocol (Table 2), screening modalities change depending on the sex and age of the patient (e.g., high ACC risk in very young children or high breast cancer risk in young women age 20–40 years).

Should surveillance be adjusted on the basis of the gene mutation (genotype) and its perceived spectrum of disease (phenotype)? Phenotype–genotype correlations may become increasingly important for risk-adapted surveillance for LFS patients. The panel is aware that there is evidence for a genotype–phenotype correlation with dominant-negative missense mutations affecting the DNA-binding domain leading to a more aggressive, early-onset phenotype, and other types of mutations being associated with a less penetrant later onset disease (73). However, the group consensus is that it is currently premature to make adjustments to the surveillance protocol based on genotype because of the lack of precise predictions for individual patients. More data from functional assays to measure the consequence of a given mutation, the presence and role of genetic modifiers, as well as clinical (registry) data will be necessary to incorporate new genotype–phenotype data as they are reported and validated. The expert panel recommends that these surveillance recommendations be reevaluated regularly as this new information becomes available, as they might be stratified in the future according to the type of mutation, family history, and other modifiers.

Conclusions

LFS is associated with a high lifelong cancer risk. It has been shown that *TP53* mutation carriers enrolled in a surveillance

program have an improved survival. Therefore, our international panel recommends that all individuals with LFS (as defined by the identification of a pathologic *TP53* germline mutation and/or by meeting the classic clinical LFS criteria) be offered surveillance as soon as the diagnosis of LFS is established. The expert panel recommends the use of the Toronto protocol with modifications, as outlined in Table 2, while being aware of the notion that not all patients will have access to medical systems offering this type of surveillance. Although the suggested surveillance strategy focuses on early cancer detection, future research studies will need to further address psychosocial impacts of such surveillance on LFS patients and possibly the development of newer molecularly based technologies for even earlier detection of the diverse malignancies. Additional data will be needed to validate and refine the surveillance strategies that comprise the Toronto protocol. In addition to simple measures, such as sun protection and avoidance of tobacco products, cancer prevention strategies will need to be explored in this high-risk condition. Notably, individuals with a germline *TP53* mutation who smoke cigarettes have been shown to be at significantly higher risk of developing lung cancer than individuals with a germline *TP53* pathogenic variant who do not smoke (78). There are no data indicating that the cancer risk is increased through global flying; however, medical radiation exposure should be limited to those investigations that are required for important treatment decisions. Finally, future research may allow us to design genotype-adopted surveillance strategies, because cancer risk may be influenced by mutation type

and genetic modifiers. However, it is too early to make precise predictions. Because of the rarity of LFS, we recommend that surveillance should be led by physicians with experience in cancer predisposition. This is also true for the radiologists interpreting the imaging studies. Shared care strategies may also be feasible. To ensure a continuous learning curve, we highly encourage the enrollment of LFS patients in national or international cancer predisposition registries that collect medical and biochemical information, electronic images, and biospecimens (blood and tissue).

Disclosure of Potential Conflicts of Interest

Grant Support

References

1. Malkin D. Li-Fraumeni syndrome. Genes Cancer 2011;2:475–84.
2. Kamihara J, Rana HQ, Garber JE. Germline TP53 mutations and the changing landscape of Li-Fraumeni syndrome. Hum Mutat 2014; 35:654–62.
3. Li FP, Fraumeni JF Jr. Soft-tissue sarcomas, breast cancer, and other neoplasms. A familial syndrome? Ann Intern Med 1969;71:747–52.
4. Li FP, Fraumeni JF Jr, Mulvihill JJ, Blattner WA, Dreyfus MG, Tucker MA, et al. A cancer family syndrome in twenty-four kindreds. Cancer Res 1988; 48:5358–62.
5. Malkin D, Li FP, Strong LC, Fraumeni JF Jr, Nelson CE, Kim DH, et al. Germ line p53 mutations in a familial syndrome of breast cancer, sarcomas, and other neoplasms. Science 1990;250:1233–8.
6. Srivastava S, Zou ZQ, Pirollo K, Blattner W, Chang EH. Germ-line transmission of a mutated p53 gene in a cancer-prone family with Li-Fraumeni syndrome. Nature 1990;348:747–9.
7. Evans DG, Birch JM, Narod SA. Is CHEK2 a cause of the Li-Fraumeni syndrome? J Med Genet 2008;45:63–4.
8. Chompret A, Abel A, Stoppa-Lyonnet D, Brugieres L, Pages S, Feunteun J, et al. Sensitivity and predictive value of criteria for p53 germline mutation screening. J Med Genet 2001;38:43–7.
9. Ruijs MW, Verhoef S, Rookus MA, Pruntel R, van der Hout AH, Hogervorst FB, et al. TP53 germline mutation testing in 180 families suspected of Li-Fraumeni syndrome: mutation detection rate and relative frequency of cancers in different familial phenotypes. J Med Genet 2010;47:421–8.
10. Gonzalez KD, Noltner KA, Buzin CH, Gu D, Wen-Fong CY, Nguyen VQ, et al. Beyond Li Fraumeni Syndrome: clinical characteristics of families with p53 germline mutations. J Clin Oncol 2009;27:1250–6.
11. Bougeard G, Renaux-Petel M, Flaman JM, Charbonnier C, Fermey P, Belotti M, et al. Revisiting Li-Fraumeni Syndrome From TP53 Mutation Carriers. J Clin Oncol 2015;33:2345–52.
12. Hettmer S, Archer NM, Somers GR, Novokmet A, Wagers AJ, Diller L, et al. Anaplastic rhabdomyosarcoma in TP53 germline mutation carriers. Cancer 2014;120:1068–75.
13. Wasserman JD, Novokmet A, Eichler-Jonsson C, Ribeiro RC, Rodriguez-Galindo C, Zambetti GP, et al. Prevalence and functional consequence of TP53 mutations in pediatric adrenocortical carcinoma: a children's oncology group study. J Clin Oncol 2015;33:602–9.
14. Holmfeldt L, Wei L, Diaz-Flores E, Walsh M, Zhang J, Ding L, et al. The genomic landscape of hypodiploid acute lymphoblastic leukemia. Nat Genet 2013;45:242–52.
15. Zhukova N, Ramaswamy V, Remke M, Pfaff E, Shih DJ, Martin DC, et al. Subgroup-specific prognostic implications of TP53 mutation in medulloblastoma. J Clin Oncol 2013;31:2927–35.
16. Kool M, Jones DT, Jager N, Northcott PA, Pugh TJ, Hovestadt V, et al. Genome sequencing of SHH medulloblastoma predicts genotype-related response to smoothened inhibition. Cancer Cell 2014;25:393–405.
17. McIntyre JF, Smith-Sorensen B, Friend SH, Kassell J, Borresen AL, Yan YX, et al. Germline mutations of the p53 tumor suppressor gene in children with osteosarcoma. J Clin Oncol 1994;12:925–30.
18. Mirabello L, Yeager M, Mai PL, Gastier-Foster JM, Gorlick R, Khanna C, et al. Germline TP53 variants and susceptibility to osteosarcoma. J Natl Cancer Inst 2015;107:djv101.
19. Zhang J, Walsh MF, Wu G, Edmonson MN, Gruber TA, Easton J, et al. Germline mutations in predisposition genes in pediatric cancer. N Engl J Med 2015;373:2336–46.
20. Hof J, Krentz S, van Schewick C, Korner G, Shalapour S, Rhein P, et al. Mutations and deletions of the TP53 gene predict nonresponse to treatment and poor outcome in first relapse of childhood acute lymphoblastic leukemia. J Clin Oncol 2011;29:3185–93.
21. Diskin SJ, Capasso M, Diamond M, Oldridge DA, Conkrite K, Bosse KR, et al. Rare variants in TP53 and susceptibility to neuroblastoma. J Natl Cancer Inst 2014;106:dju047.
22. Rana HQ, Gelman R, Thompson J, McFarland R, LaDuca H, Dalton E, et al. Single gene (SG) vs. multi-gene panel (MGP) testing for TP53 germline mutations in Li Fraumeni syndrome (LFS) [abstract]. In: Proceedings of the 2015 American Society of Human Genetics; 2015 Oct 6–10; Baltimore, MD. Bethesda, MD: ASHG; 2015. Abstract nr PgmNr2667.
23. Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SA, Behjati S, Biankin AV, et al. Signatures of mutational processes in human cancer. Nature 2013;500:415–21.

24. Rausch T, Jones DT, Zapatka M, Stutz AM, Zichner T, Weischenfeldt J, et al. Genome sequencing of pediatric medulloblastoma links catastrophic DNA rearrangements with TP53 mutations. Cell 2012;148:59–71.

25. Varley JM, McGown G, Thorncroft M, James LA, Margison GP, Forster G, et al. Are there low-penetrance TP53 Alleles? evidence from childhood adrenocortical tumors. Am J Hum Genet 1999;65:995–1006.

26. Lalloo F, Varley J, Moran A, Ellis D, O'Dair L, Pharoah P, et al. BRCA1, BRCA2 and TP53 mutations in very early-onset breast cancer with associated risks to relatives. Eur J Cancer 2006;42:1143–50.

27. Mouchawar J, Korch C, Byers T, Pitts TM, Li E, McCredie MR, et al. Population-based estimate of the contribution of TP53 mutations to subgroups of early-onset breast cancer: australian breast cancer family study. Cancer Res 2010;70:4795–800.

28. Nichols KE, Malkin D. genotype versus phenotype: the yin and yang of germline TP53 mutations in li-fraumeni syndrome. J Clin Oncol 2015; 33:2331–3.

29. Malkin D, Jolly KW, Barbier N, Look AT, Friend SH, Gebhardt MC, et al. Germline mutations of the p53 tumor-suppressor gene in children and young adults with second malignant neoplasms. N Engl J Med 1992;326:1309–15.

30. Hisada M, Garber JE, Fung CY, Fraumeni JF Jr, Li FP. Multiple primary cancers in families with Li-Fraumeni syndrome. J Natl Cancer Inst 1998; 90:606–11.

31. Nutting C, Camplejohn RS, Gilchrist R, Tait D, Blake P, Knee G, et al. A patient with 17 primary tumours and a germ line mutation in TP53: tumour induction by adjuvant therapy? Clin Oncol (R Coll Radiol) 2000;12:300–4.

32. Limacher JM, Frebourg T, Natarajan-Ame S, Bergerat JP. Two metachronous tumors in the radiotherapy fields of a patient with Li-Fraumeni syndrome. Int J Cancer 2001;96:238–42.

33. Izawa N, Matsumoto S, Manabe J, Tanizawa T, Hoshi M, Shigemitsu T, et al. A Japanese patient with Li-Fraumeni syndrome who had nine primary malignancies associated with a germline mutation of the p53 tumor-suppressor gene. Int J Clin Oncol 2008;13:78–82.

34. Heymann S, Delaloge S, Rahal A, Caron O, Frebourg T, Barreau L, et al. Radio-induced malignancies after breast cancer postoperative radiotherapy in patients with Li-Fraumeni syndrome. Radiat Oncol 2010;5:104.

35. Mai PL, Best AF, Peters JA, DeCastro RM, Khincha PP, Loud JT, et al. Risks of first and subsequent cancers among TP53 mutation carriers in the National Cancer Institute Li-Fraumeni syndrome cohort. Cancer 2016;122:3673–81.

36. Ariffin H, Hainaut P, Puzio-Kuter A, Choong SS, Chan AS, Tolkunov D, et al. Whole-genome sequencing analysis of phenotypic heterogeneity and anticipation in Li-Fraumeni cancer predisposition syndrome. Proc Natl Acad Sci U S A 2014;111:15497–501.

37. Mai PL, Malkin D, Garber JE, Schiffman JD, Weitzel JN, Strong LC, et al. Li-Fraumeni syndrome: report of a clinical research workshop and creation of a research consortium. Cancer Genet 2012;205:479–87.

38. Garritano S, Gemignani F, Palmero EI, Olivier M, Martel-Planche G, Le Calvez-Kelm F, et al. Detailed haplotype analysis at the TP53 locus in p.R337H mutation carriers in the population of Southern Brazil: evidence for a founder effect. Hum Mutat 2010;31:143–50.

39. Achatz MI, Olivier M, Le Calvez F, Martel-Planche G, Lopes A, Rossi BM, et al. The TP53 mutation, R337H, is associated with Li-Fraumeni and Li-Fraumeni-like syndromes in Brazilian families. Cancer Lett 2007; 245:96–102.

40. Achatz MI, Zambetti GP. The inherited p53 mutation in the Brazilian population. Cold Spring Harb Perspect Med 2016;6:pii:a026195.

41. Aubrey BJ, Strasser A, Kelly GL. Tumor-suppressor functions of the TP53 pathway. Cold Spring Harb Perspect Med 2016;6:pii:a026104.

42. Olivier M, Goldgar DE, Sodha N, Ohgaki H, Kleihues P, Hainaut P, et al. Li-Fraumeni and related syndromes: correlation between tumor type, family structure, and TP53 genotype. Cancer Res 2003;63:6643–50.

43. Chompret A, Brugieres L, Ronsin M, Gardes M, Dessarps-Freichey F, Abel A, et al. P53 germline mutations in childhood cancers and cancer risk for carrier individuals. Br J Cancer 2000;82:1932–7.

44. Bond GL, Hu W, Bond EE, Robins H, Lutzker SG, Arva NC, et al. A single nucleotide polymorphism in the MDM2 promoter attenuates the p53 tumor suppressor pathway and accelerates tumor formation in humans. Cell 2004;119:591–602.

45. Sagne C, Marcel V, Bota M, Martel-Planche G, Nobrega A, Palmero EI, et al. Age at cancer onset in germline TP53 mutation carriers: association with

46. Gemignani F, Moreno V, Landi S, Moullan N, Chabrier A, Gutierrez-Enriquez S, et al. A TP53 polymorphism is associated with increased risk of colorectal cancer and with reduced levels of TP53 mRNA. Oncogene 2004;23:1954–6.

47. Tabori U, Nanda S, Druker H, Lees J, Malkin D. Younger age of cancer initiation is associated with shorter telomere length in Li-Fraumeni syndrome. Cancer Res 2007;67:1415–8.

48. Id Said B, Kim H, Tran J, Novokmet A, Malkin D. Super-transactivation TP53 variant in the germline of a family with li-fraumeni syndrome. Hum Mutat 2016;37:889–92.

49. Samuel N, Wilson G, Lemire M, Id Said B, Lou Y, Li W, et al. Genome-wide DNA methylation analysis reveals epigenetic dysregulation of microRNA-34A in TP53-associated cancer susceptibility. J Clin Oncol 2016 Aug 22. [Epub ahead of print].

50. Samuel N, Wilson G, Id Said B, Pan A, Deblois G, Fischer NW, et al. Transcriptome-wide characterization of the endogenous miR-34A-p53 tumor suppressor network. Oncotarget 2016;7:49611–22.

51. Shlien A, Tabori U, Marshall CR, Pienkowska M, Feuk L, Novokmet A, et al. Excessive genomic DNA copy number variation in the Li-Fraumeni cancer predisposition syndrome. Proc Natl Acad Sci U S A 2008;105:11264–9.

52. Ballinger ML, Mitchell G, Thomas DM. Surveillance recommendations for patients with germline TP53 mutations. Curr Opin Oncol 2015;27: 332–7.

53. McBride KA, Ballinger ML, Killick E, Kirk J, Tattersall MH, Eeles RA, et al. Li-Fraumeni syndrome: cancer risk assessment and clinical management. Nat Rev Clin Oncol 2014;11:260–71.

54. Daly MB, Pilarski R, Berry M, Buys SS, Farmer M, Friedman DVM, et al. NCCN clinical practical guidelines in oncology genetic/familial high-risk assessment: breast and ovarian. Fort Washington, PA: National Comprehensive Cancer Network; 2017. Available from: https://www.nccn.org/professionals/physician_gls/f_guidelines.asp#genetics_screening.

55. Villani A, Tabori U, Schiffman J, Shlien A, Beyene J, Druker H, et al. Biochemical and imaging surveillance in germline TP53 mutation carriers with Li-Fraumeni syndrome: a prospective observational study. Lancet Oncol 2011;12:559–67.

56. Villani A, Shore A, Wasserman JD, Stephens D, Kim RH, Druker H, et al. Biochemical and imaging surveillance in germline TP53 mutation carriers with Li-Fraumeni syndrome: 11 year follow-up of a prospective observational study. Lancet Oncol 2016;17:1295–305.

57. Greer M-LC, Voss SD, States LJ. Pediatric cancer predisposition imaging: focus on whole-body MRI. Clin Cancer Res 2017;23:e6–e13.

58. Masciari S, Van den Abbeele AD, Diller LR, Rastarhuyeva I, Yap J, Schneider K, et al. F18-fluorodeoxyglucose-positron emission tomography/computed tomography screening in Li-Fraumeni syndrome. JAMA 2008;299: 1315–9.

59. Nogueira ST, Lima EN, Nobrega AF, Torres Ido C, Cavicchioli M, Hainaut P, et al. (18)F-FDG PET-CT for surveillance of brazilian patients with li-fraumeni syndrome. Front Oncol 2015;5:38.

60. Custodio G, Parise GA, Kiesel Filho N, Komechen H, Sabbaga CC, Rosati R, et al. Impact of neonatal screening and surveillance for the TP53 R337H mutation on early detection of childhood adrenocortical tumors. J Clin Oncol 2013;31:2619–26.

61. Peterson SK, Pentz RD, Marani SK, Ward PA, Blanco AM, LaRue D, et al. Psychological functioning in persons considering genetic counseling and testing for Li-Fraumeni syndrome. Psychooncology 2008;17:783–9.

62. Lammens CR, Aaronson NK, Wagner A, Sijmons RH, Ausems MG, Vriends AH, et al. Genetic testing in Li-Fraumeni syndrome: uptake and psychosocial consequences. J Clin Oncol 2010;28:3008–14.

63. Lammens CR, Bleiker EM, Aaronson NK, Wagner A, Sijmons RH, Ausems MG, et al. Regular surveillance for Li-Fraumeni Syndrome: advice, adherence and perceived benefits. Familial Cancer 2010;9:647–54.

64. Peters JA, Kenen R, Bremer R, Givens S, Savage SA, Mai PL. Easing the burden: describing the role of social, emotional and spiritual support in research families with li-fraumeni syndrome. J Genet Counseling 2016;25:529–42.

65. Lammens CR, Bleiker EM, Verhoef S, Ausems MG, Majoor-Krakauer D, Sijmons RH, et al. Distress in partners of individuals diagnosed with or at high risk of developing tumors due to rare hereditary cancer syndromes. Psychooncology 2011;20:631–8.

66. Wakefield CE, Hanlon LV, Tucker KM, Patenaude AF, Signorelli C, McLoone JK, et al. The psychological impact of genetic information on children: a systematic review. Genet Med 2016;18:755–62.
67. Gopie JP, Vasen HF, Tibben A. Surveillance for hereditary cancer: does the benefit outweigh the psychological burden?–A systematic review. Crit Rev Oncol Hematol 2012;83:329–40.
68. Townsend A, Adam S, Birch PH, Lohn Z, Rousseau F, Friedman JM. "I want to know what's in Pandora's Box": comparing stakeholder perspectives on incidental findings in clinical whole genomic sequencing. Am J Med Genet A 2012;158A:2519–25.
69. Patenaude AF, Schneider KA, Kieffer SA, Calzone KA, Stopfer JW, Basili LA, et al. Acceptance of invitations for p53 and BRCA1 predisposition testing: Factors influencing potential utilization of cancer genetic testing. Psychooncology 1996;5:241–50.
70. Oppenheim D, Brugieres L, Chompret A, Hartmann O. The psychological burden inflicted by multiple cancers in Li-fraumeni families: five case studies. J Gen Counsel 2001;10:169–83.
71. Alderfer MA, Zelley K, Lindell RB, Novokmet A, Mai PL, Garber JE, et al. Parent decision-making around the genetic testing of children for germline TP53 mutations. Cancer 2015;121:286–93.
72. Druker H, Zelley K, McGee RB, Scollon S, Kohlmann W, Schneider KA, et al. Genetic counseling for cancer predisposition in children. Clin Cancer Res 2017;23: doi: 10.1158/1078-0432.CCR-17-0834.
73. Zerdoumi Y, Aury-Landas J, Bonaiti-Pellie C, Derambure C, Sesboue R, Renaux-Petel M, et al. Drastic effect of germline TP53 missense mutations in Li-Fraumeni patients. Hum Mutat 2013;34:453–61.
74. Kanda T, Ishii K, Kawaguchi H, Kitajima K, Takenaka D. High signal intensity in the dentate nucleus and globus pallidus on unenhanced T1-weighted MR images: relationship with increasing cumulative dose of a gadolinium-based contrast material. Radiology 2014;270:834–41.
75. Flood TF, Stence NV, Maloney JA, Mirsky DM. Pediatric brain: repeated exposure to linear gadolinium-based contrast material is associated with increased signal intensity at unenhanced T1-weighted MR imaging. Radiology 2017;282:222–8.
76. Hu HH, Pokorney A, Towbin RB, Miller JH. Increased signal intensities in the dentate nucleus and globus pallidus on unenhanced T1-weighted images: evidence in children undergoing multiple gadolinium MRI exams. Pediatr Radiol 2016;46:1590–8.
77. Porter CC, Druley TE, Erez A, Kuiper RP, Onel K, Schiffman JD, et al. Recommendations for surveillance for children with leukemia-predisposing conditions. Clin Cancer Res 2017;23:e14–e22.
78. Hwang SJ, Cheng LS, Lozano G, Amos CI, Gu X, Strong LC. Lung cancer risk in germline p53 mutation carriers: association between an inherited cancer predisposition, cigarette smoking, and cancer risk. Human Genet 2003;113:238–43.

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